

The Immunomodulatory and Metabolic Effects of *Chlorella Vulgaris* Biologically Active Compounds on Experimental Models and Bovine Physiology

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Abstract This study investigates the efficacy of biologically active substances (BAS) derived from *Chlorella vulgaris* in correcting secondary immunodeficiency induced by toxic hepatitis (CCl₄ and D-galactosamine) in laboratory rats and its metabolic impact on calves. Results indicated that a 1.0% concentration of *Chlorella* BAS significantly restored antibody-forming cells (AFC) in the spleen, comparable to the reference drug Thymalin. In calves, parenteral administration improved T and B-lymphocyte counts and increased body weight gain (up to 62 kg). The findings suggest that *Chlorella vulgaris* acts as a potent biostimulator of hematopoiesis and protein-carbohydrate metabolism.

Keywords *C. vulgaris*, Bioactive compounds, Immunomodulation, Toxic hepatitis, Bovine physiology, Hematopoiesis, Metabolic status

1. Introduction

The increasing impact of anthropogenic factors and xenobiotics has led to widespread physiological disruptions in livestock, particularly affecting liver function and immune response. Toxic hepatitis often results in secondary immunodeficiency, reducing animal productivity and increasing susceptibility to infectious diseases. [3]. While synthetic drugs are commonly used, their side effects necessitate the search for natural alternatives. [13]. *Chlorella vulgaris* is a unicellular green alga rich in proteins (50–55%), vitamins (A, B-group, C, E), and essential minerals. [1]. Its unique "Chlorella Growth Factor" (CGF) and polysaccharides exhibit potent antioxidant and immunomodulatory properties. [2, 11]. This study aims to scientifically substantiate the use of local *Chlorella vulgaris* strains as biostimulators for restoring physiological and immunological status.

2. Materials and Methods

2.1. Experimental Design

The research was conducted on outbred white mice (18–22 g), white rats (180–200 g), and young calves. Toxic hepatitis was induced in mice using a CCl₄ oil solution (0.2 mg/kg, i.p. for 3 days) and in rats using D-galactosamine

hydrochloride (1.1 g/kg, i.p.). [15].

2.2. Immunological and Biochemical Analysis

Antibody-forming cells (AFC) in the spleen were determined using the local hemolysis method in agarose (Jerne and Nordin). Blood parameters (erythrocytes, leukocytes) were analyzed using a Goryayev chamber. Biochemical markers (ALT, AST, protein fractions, glucose, and minerals) were measured in blood serum using standard diagnostic kits. [23].

3. Results and Discussion

3.1. Impact on Experimental Immunodeficiency

Induction of hepatitis led to a 2.7-fold decrease in AFC production in the spleen ($5,334 \pm 725$ vs. $14,621 \pm 1,029$ in intact mice). Administration of *Chlorella vulgaris* BAS resulted in a significant recovery, increasing AFC counts by 2.5 to 6.1 times depending on the concentration (Table 1). [8,17].

3.2. Physiological Status in Calves

Parenteral administration of *Chlorella* to calves stimulated T-lymphocyte (increase of 1.3–3.1%) and B-lymphocyte (1.5% increase) populations. [14,18].

Significant improvements were noted in protein metabolism: albumin levels reached 33.13 ± 0.34 g/l in the experimental group compared to the control ($P < 0.05$). Glucose levels increased by 12.3%, indicating enhanced energy homeostasis (Table 2).

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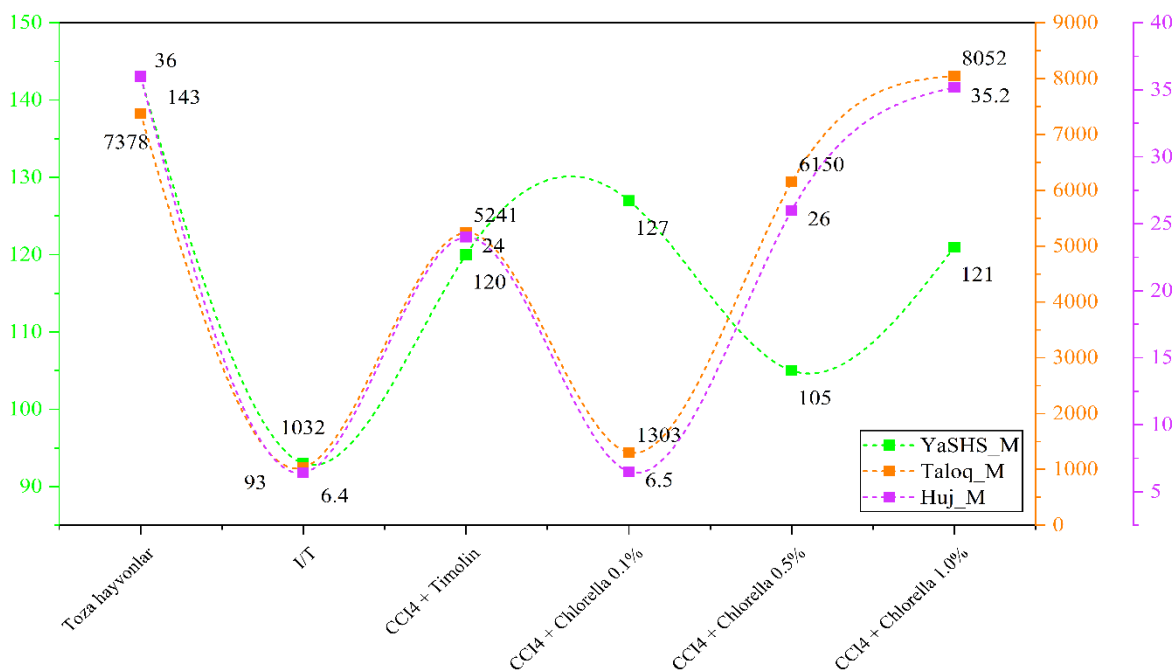
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Table 1. Comparative activity of *Chlorella vulgaris* on AFC production in experimental immunodeficiency

No.	Experimental groups	Nucleated cells ($\times 10^6$)		Plasma cell			
				AFC per Spleen.		ASCs per 10^6 splenocytes	
		M \pm m	i/n	M \pm m	i/n	M \pm m	i/n
1.	Intact group	143 \pm 16	-	7378 \pm 503	-	36,0 \pm 4,0	-
2.	Immuno-Deficiency	93 \pm 0,8	-1,6	1032 \pm 203*	-6,1	6,4 \pm 1,5*	-5,6
3.	I/D+ Timalin	120 \pm 13	+1,3	5241 \pm 637*	+5,0	24,0 \pm 2,1*	+3,7
4.	I/D+ <i>Chlorella vulgaris</i> 1%	121 \pm 12	+1,3	8052 \pm 673**	+6,1	35,2 \pm 3,7**	+5,5

Note: * p < 0.05 compared to intact; ** p < 0.05 compared to CCl₄ control

**Figure 1****Table 2.** Protein and carbohydrate metabolism indicators

Indicators	Groups		Physiological norm
	Control	Experience	
Total protein (g,l)	86,2 \pm 3,78	80,5 \pm 2,47	70-92
Albumin (g,l)	30,7 \pm 0,63	33,13 \pm 0,34*	25-36
Globulin (g,l)	55,5 \pm 4,12	47,4 \pm 2,78	40-67
Urea, mol/l	3,8 \pm 0,16	5,34 \pm 0,24*	2,35-7,06
ALT, ME/l	34,6 \pm 5,10	33,3 \pm 2,31	12-35
ACT, ME/l	73,0 \pm 7,54	106,2 \pm 11,17	46-108
Creatinine, mkmol/l	77,2 \pm 3,53	93,3 \pm 5,97	63-162
Glucose, mmol/l	3,7 \pm 0,11	4,1 \pm 0,03*	1,65-4,19

Note: * — P<0.05; + — P<0.10

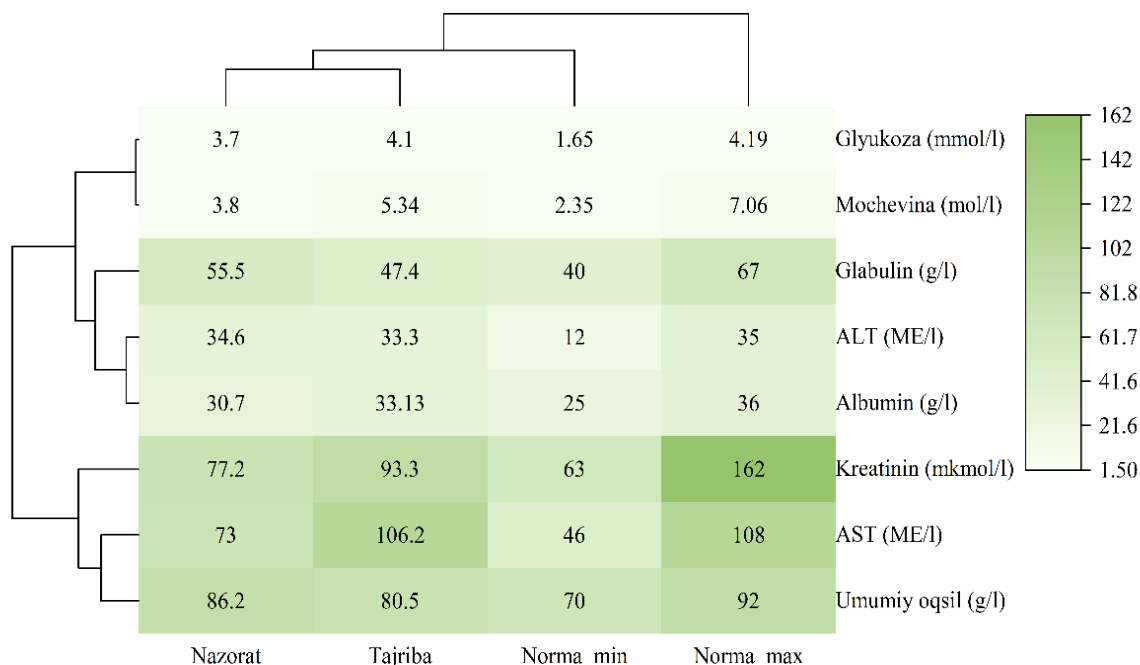


Figure 2. Dynamics of protein and carbohydrate metabolism under the influence of the drug

4. Analysis of Protein and Carbohydrate Metabolism

Biochemical profiling of blood serum demonstrated a distinct predominance of anabolic processes in the experimental group administered with *Chlorella vulgaris* suspension. Specifically, serum albumin levels in the experimental group exhibited a statistically significant increase ($P < 0.05$) compared to the control, reaching 33.13 ± 0.34 g/L. This elevation in albumin suggests a robust stimulation of the liver's protein-synthetic function. This physiological shift is attributed to the direct systemic bioavailability of the preparation's biologically active compounds, specifically bioactive peptides and essential amino acids, which facilitate enhanced hepatic proteosynthesis and optimize molecular transport mechanisms. [24].

The protein index (albumin/globulin ratio) reached 0.68 in the experimental cohort, whereas it remained at 0.58 in the control group. This divergence indicates an improved potential for replenishing systemic protein reserves and a superior level of cellular supply with essential plastic materials. [22].

Furthermore, a positive trajectory was observed in energy metabolism markers. Glucose concentrations in the experimental group increased by 12.3% (0.45 mmol/L) relative to the control ($P < 0.05$), signifying an enhanced energetic supply to peripheral tissues and improved glucose homeostasis.

The rise in creatinine levels from $77.2 \mu\text{mol/L}$ (control) to $93.3 \mu\text{mol/L}$ (experimental) indicates an accelerated metabolic turnover within muscle tissues under the influence of the biostimulant. [22]. This correlates with an expansion of the phosphocreatine pool, a vital energy accumulator. Such an increase within the physiological reference range is of paramount importance for ensuring efficient ATP resynthesis

and maintaining high stability in intracellular bioenergetics.

5. Conclusions

The comprehensive physiological and immunological assessment of *Chlorella vulgaris* biologically active substances (BAS) allows for the following conclusions:

1. Potent Immunomodulatory Efficacy: It was established that *Chlorella vulgaris* biomass serves as a high-efficiency biostimulant for correcting secondary immunodeficiency induced by hepatotropic toxins (CCl_4 and D-galactosamine). The administration of a 1.0% suspension demonstrated a superior restorative effect on the splenic immune microenvironment, increasing the population of antibody-forming cells (AFC) by 6.1 times, effectively neutralizing the immunosuppressive effects of toxic hepatitis.
2. Hematopoietic and Regenerative Stimulation: The preparation exhibits a pronounced hematopoietic effect, specifically stimulating erythropoiesis and leucopoiesis within the bone marrow. This leads to a stabilized balance of peripheral blood formed elements, ensuring systemic homeostasis without inducing pathological leucocytosis.
3. Metabolic Optimization in Livestock: Parenteral administration of *Chlorella* BAS in calves and cows significantly enhances anabolic processes. The observed increase in serum albumin (33.13 ± 0.34 g/L) and glucose levels (12.3%) indicates an optimization of hepatic proteosynthesis and energy metabolism. Furthermore, the stabilization of the creatinine-phosphocreatine pool confirms improved intracellular

bioenergetics and ATP resynthesis capacity.

4. Practical Applicability: Given its high biological activity, cost-effectiveness, and absence of adverse side effects compared to synthetic analogs (e.g., Thymalin), the locally derived *Chlorella vulgaris* strain is a viable candidate for integration into veterinary pharmacology. Its use is recommended for enhancing the nonspecific resistance and productive potential of agricultural animals, particularly in ecologically unfavorable regions.

In summary, this research scientifically substantiates *Chlorella vulgaris* as a multifaceted biostimulant that effectively bridges the gap between hepatoprotection and immunocorrection in animal physiology.

REFERENCES

- [1] Mendes, A. R., Spíndola, M. P., Lordelo, M., & Prates, J. A. M. (2024). Chemical Compounds, Bioactivities, and Applications of *Chlorella vulgaris* in Food, Feed and Medicine. *Applied Sciences*, 14(23), 10810. <https://doi.org/10.3390/app142310810>.
- [2] Bito, T., Okumura, E., Fujishima, M., & Watanabe, F. (2020). Potential of *Chlorella* as a Dietary Supplement to Promote Human Health. *Nutrients*, 12(9), 2524. <https://doi.org/10.3390/nu12092524>.
- [3] Panahi, Y., Mostafazadeh, B., Abrishami, A., et al. (2013). Investigation of the effects of *Chlorella vulgaris* supplementation on the modulation of oxidative stress in apparently healthy smokers. *Clinical Laboratory*, 59(5-6), 579-587.
- [4] Gouveia, L., Batista, A. P., Sousa, I., Raymundo, A., & Bandarra, N. M. (2018). Microalgae in Novel Food Products. In *Microalgae as a Source of Biochemicals and Functional Ingredients* (pp. 1-65). Taylor & Francis.
- [5] Martins, C. F., Pestana, J. M., Alfaia, C. M., et al. (2021). Effects of *Chlorella vulgaris* as a feed ingredient on the quality and nutritional value of weaned piglets' meat. *Foods*, 10(5), 1155. <https://doi.org/10.3390/foods10051155>.
- [6] Abdel-Tawwab, M., Mousa, M. A., & Mamoon, A. (2022). Dietary *Chlorella vulgaris* modulates the performance, antioxidant capacity, innate immunity, and disease resistance capability of Nile tilapia fed on plant-based diets. *Animal Feed Science and Technology*, 283, 115181.
- [7] Latif, A. A., Assar, D. H., Hamza, H. A., et al. (2021). Protective role of *Chlorella vulgaris* against paracetamol-induced toxic effects on haematological, biochemical, and oxidative stress parameters in Wistar rats. *Scientific Reports*, 11, 3911. <https://doi.org/10.1038/s41598-021-83445-w>.
- [8] Queiroz, M. L., da Rocha, M. C., Torello, C. O., et al. (2011). *Chlorella vulgaris* restores bone marrow cellularity and cytokine production in lead-exposed mice. *Food and Chemical Toxicology*, 49(11), 2934-2941.
- [9] Safi, C., Zebib, B., Merah, O., et al. (2014). Morphology, composition, production, processing and applications of *Chlorella vulgaris*: A review. *Renewable and Sustainable Energy Reviews*, 35, 265-278.
- [10] Tsiplakou, E., Abdullah, M., Mavrommatis, A., et al. (2017). The effect of dietary *Chlorella vulgaris* inclusion on goat's milk chemical composition and fatty acids profile. *Journal of Animal Physiology and Animal Nutrition*, 102(1), 142-151.
- [11] Buono, S., et al. (2014). Biological activities of chlorophyll and carotenoids from *Chlorella vulgaris*. *Food Chemistry*, 165, 33-40.
- [12] Butler, W. R. (2003). Energy balance relationships with follicular development, ovulation and fertility in postpartum dairy cows. *Livestock Production Science*, 83(2-3), 211-218.
- [13] Chang, S. H., et al. (2020). Natural versus synthetic immunomodulators: Efficacy and safety profiles. *Biomedicine & Pharmacotherapy*, 128, 110268.
- [14] El-Bahr, S. M., et al. (2020). Effect of Microalgae on Antioxidant Status and immune response in Calves. *Veterinary World*, 13(9), 1962-1969.
- [15] El-Baky, H. H. A., & El-Baroty, G. S. (2009). Hepatoprotective effect of *Chlorella vulgaris* extract against liver toxicity. *Journal of Basic Microbiology*, 49(S1), S20-S27.
- [16] Gimpl, G., & Fahrenholz, F. (2001). The oxytocin receptor system: structure, function, and regulation. *Physiological Reviews*, 81(2), 629-683.
- [17] Hasegawa, T., et al. (1997). Effect of hot water extract of *Chlorella vulgaris* on cytokine production. *International Journal of Immunopharmacology*, 19(6), 312-320.
- [18] Kholif, A. E., et al. (2017). Effect of feeding *Chlorella vulgaris* on performance, milk quality and serum biochemical parameters of lactating goats/cows. *Animal Feed Science and Technology*, 223, 102-114.
- [19] LeBlanc, S. J. (2008). Postpartum uterine disease and dairy herd reproductive performance: A review. *The Veterinary Journal*, 176(1), 102-114.
- [20] Pan, L., et al. (2023). Post-pandemic immune health strategies: The role of natural compounds. *Journal of Clinical Medicine*, 12(4), 1120.
- [21] Rozenberg, S., et al. (2022). Algae as a functional feed supplement for cattle: Immune modulation and stress mitigation. *Frontiers in Veterinary Science*, 9, 876321.
- [22] Saeid, A., & Chameh, H. (2019). Microalgae (*Chlorella vulgaris*) as a Feed Supplement for Dairy Cattle: Effects on Milk Yield and Composition. *Journal of Dairy Science*, 102(7), 6120-6130.
- [23] Sheldon, I. M., & Dobson, H. (2004). Postpartum uterine health in cattle. *Animal Reproduction Science*, 82-83, 295-306.
- [24] Wang, Y., et al. (2023). Bioactive components of microalgae and their application in health. *Foods*, 12(7), 1354.