

Facile Root for Isolation of Rhamnocitrin Sulphate from *Tetracera Alnifolia* Willd

Victor N’Goka^{1,2,3}, Nicaise Narcisse Obaya^{2,3}, Jean Bruno Mokoko^{3,4,*}, Cyril Antheaume⁵

¹Department of Chemistry, Pharmacochemical and Pharmacotechnical Laboratory of Medicinal Plants, Chired, Congo

²Faculty of Science and Technology

³Mariem Ngouabi University, Brazzaville, Congo

⁴Masters Department, Faculty of Science and Health, Pharmacology and Biochemistry Laboratory

⁵Laboratory of NMR, LC1 IFR85, ULP, Faculty of Pharmacy, Strasbourg, France

Abstract Rhamnocitrin 3-sulphate has been isolated with one percent yield, from *Tetracera alnifolia*, a non-cultivated medicinal plant from Congo. Rhamnocitrin 3-sulphate has been isolated after recrystallization of crude dried extract obtained from aqueous decoction of *Tetracera* leaves and stem. In this paper, Rhamnocitrin 3-sulphate has been easily isolated for the first time from *Tetracera alnifolia* and the corresponding tri-acetylated compound has been synthesized. Identification, characterisation and structure elucidation have been carried out using TLC, HPLC, UV-VIS, NMR and MS analyses in comparison with those described in the literature.

Keywords Rhamnocitrin 3-sulphate, Rhamnocitrin, *Tetracera alnifolia*, Dilleniaceae, Flavonol

1. Introduction

Nowadays World Health Organization (WHO) exhorts African States to produce traditional ameliorated medicine. However, the development of these products has been subordinated to the standardisation of the procedure for their elaboration. This paper describes the isolation of Rhamnocitrin 3-sulphate with high amount in one step, instead of the traditional multistep procedure of isolation of molecule from vegetable matter as shown by Yamauchi et al. and Chaabi et al. for examples [1,2]. *Tetracera alnifolia* Willd is a perennial, evergreen big liana of the family of Dilleniaceae. It is growing wildly in the forest and most warm regions of the world especially in Senegal, Central African Republic, Gabon, Cameroon, Congo, Congo RD, Angola, Fernando Po and Zambia. *T. alnifolia* leaves and stem have been traditionally used for treating venereal diseases, abdominal pains and to regulate menstruation [3,4], which indicates probably the presence of antibacterial compounds in this plant. Preliminary antibacterial study of *T. alnifolia* have been reported and showed substantial activity against *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Streptococcus faecalis*, *Echerichia coli*, *Shigella dysenteriae*, *Yersinia enterocolitica*, *Enterobacter cloacea*, *Salmonella*

sp. abony, *Serretia marcescens* and *Pseudomonas aeruginosa* [5]. Nowadays *T. alnifolia* is used in association with other medicinal plants to make traditional ameliorated medicine sold in Congolese pharmacies; but the chemical composition of this antimicrobial and anti-inflammatory preparation is unknown. In our knowledge no chemical composition of *T. alnifolia* extract has been reported. Rhamnocitrin, a 7- methyl-Kaempferol (Figure 1), has been identified in many plants [6-10] and this interesting compound has been isolated as the corresponding triacetoxy derivative in little amount, 35 mg by Rossi et al. [8]. Rhamnocitrin has been reported to show several biological activities. It has antibacterial, anti-inflammatory effect [11], including strong inhibition of platelet aggregation induced by arachidonic acid [12]. Its anti-inflammatory and antispasmodic activities have been reported [13] as well as its strong antioxidant activity [11]. Qualitative studies of flavonoid sulphates have been well documented and particularly Rhamnocitrin 3-sulphate has been identified from *Tetracera poggei* Gilg and *T. alnifolia* by Gurni et al. [14,15]. Silva and collaborators [10] described Rhamnocitrin by long and expensive usually method of extraction and isolation from *Solanum jабrense* and *S. Paludosum*. In the present work we report an easy and cheap method for quantitative isolation and purification of Rhamnocitrin 3-sulphate from *T. alnifolia* using crystallization only. The molecular formula was elucidated on the basis of spectroscopic studies in comparison with those described in the literature.

* Corresponding author:

jbmokoko@yahoo.fr (Jean Bruno Mokoko)

Published online at <http://journal.sapub.org/aac>

Copyright © 2020 The Author(s). Published by Scientific & Academic Publishing

This work is licensed under the Creative Commons Attribution International

License (CC BY). <http://creativecommons.org/licenses/by/4.0/>

2. Materials and Methods

1. Plant Material

Leaves and stem of *T. alnifolia* Willd have been collected near MASSENGO area, in the north of Brazzaville, Congo. The plant sample was kindly authenticated by comparison with a voucher specimen at the Herbarium of the "Centre de Recherche sur les Ressources Végétales (CERVE)" in Brazzaville, Congo.

2. Preparation of the crude extract from *T. alnifolia* Willd

Dried powdered, 25 g, of leaves and stem of *T. alnifolia* were boiling in 300 ml of water for 30 min with shaking. The remaining solution has been filtered using Whatman paper n° 40; 8 µm. The filtrate solution has been concentrated *in vacuo* at 40 °C still dryness to give a Rhamnocitrin 3-sulphate rich extract. And then, 250 mg of pure Rhamnocitrin 3-sulphate, has been obtained after crystallization of the above dry crude extract in water. Experience was carried out in triplicate. The yield of Rhamnocitrin 3-sulphate was of 1% w/w from dried vegetable material.

3. Experimental Equipment

NMR spectra were recorded with a Bruker Avance DPX-300 spectrometer (at 300 MHz for ¹H and 75 MHz for ¹³C NOESY spectra were obtained at 400 MHz. Mass spectra were taken on a ESI Bruker Daltonics HCT ion trap mass spectrometer. The UV spectra were obtained on a HPLC system: pump: VARIAN 9010, injector: VARIAN 9100, detector: VARIAN Prostar 330 provided with a pin of diodes. TLC Plates, Silica gel Merck 60 F254, (20x20 cm) were used. The flavonoides bands were visualized by spraying with Dragendorff's reagent, followed by 10% HCl, and or using ultraviolet light at 254 and 365 nm.

4. Results and Discussion

Pure compound was isolated as a yellow amorphous powder, and the purity was confirmed by TLC, R_f = 0.46 (Ethyl acetate-methanol: 8-2); HPLC and SM (Figure 1 and 2). The structure (Figure 3) was elucidated by comparison of the spectroscopic data with the corresponding literature values for the un-sulphated compounds [9,10,17-21]. The ESI-MS and EI-MS have been analysed by comparison of the previously described for Kaempferol [20]. The product ion mass spectrum for Rhamnocitrin 3-sulphate m/z 380 is shown in figure 2. A peak at m/z 379 [M-H] was consistent with a molecular formula C₁₆H₁₂SO₉-H; m/z 315 [M-H-SO₂] corresponding to the loss HSO₂; m/z 299 [M-HSO₃] corresponding to the loss of HSO₃; m/z 284 [M-HSO₃-CH₃] corresponding to the demethylated of later product; m/z 271 [M-HSO₃-C=O] corresponding to the loss of HSO₃ and C=O; m/z 255 [M-HSO₃-CH₃-HC=O] corresponding to the loss of HSO₃, HC=O and demethylation; m/z 243

[M-HSO₃-CH₃-CH=C=O] corresponding to the loss of HSO₃, CH=C=O and demethylation; m/z 227 (M-HSO₃-OCH₃-CH=C=O) corresponding to the loss of HSO₃, CH=C=O and demethoxylation.

The ¹³C NMR and DEPT spectrum (Figure 4) showed signals due to 16 carbons comprising one methoxyl, six aromatic methine, nine quaternary carbons from where; two fully substituted aromatic carbons, two oxymethine, one bearing a carbonyl and four bearing an oxygen atom from where two similar bearing hydroxyl, one bearing sulphate and the last for the methoxyl. These fragments account for a molecular formula C₁₆H₁₂SO₉, indicating that the two remaining hydrogen atoms are present in hydroxyl groups. The ¹H NMR spectrum (Figure 5) showed signal at δ 3.86 ppm typical of methyl ether (OMe) by three proton singlet. Also in ¹³C NMR spectrum the presence of methyl ether group was brought in evidence by the signal observed at δ 56. In the ¹H-¹H NOESY (Figure 6A) contour plot the methine H-2' and H-6' showed correlations to both H-3' and H-5' which confirmed the presence of the 1', 4'- substituted phenyl fragment. The presence of a methoxyl group at C-7 as shown in figure 6B was confirmed by the nOe correlations between the proton in C-7 and the protons in C-6 and C-8 (Figure 6 and figure 7). The presence of a sulphate group at C-3 in the latter compound was confirmed by the signal found at δ 156.6 instead of δ 136 for the un-sulphated Rhamnocitrin compound. The ¹H NMR spectrum of acetylated compound showed three acetoxy group at δ 2.41, 2.32, 2.30 besides signal characteristic of methoxyl group at δ 3.89 and signals of protons bearing by carbons C-6 and C-8 became doublet with allylic J=2.4 Hz showing meta position of their protons. Comparisons between acetylated and non acetylated compound allowed the assignment of the positions of the acetoxy and methoxyl groups. In the acetylated compound, the signals of both H-2' and H-6' shift upfield for 0.71 ppm this is in accordance with the OAc in C-3 position (Rossi *et al.*, 1997), the signals for H-3' and H-5' move 0.37 ppm downfield this is in accordance with the OAc at C-4'. We observed a little diamagnetic effect for the signal of H-6 and H-8. Signals of H-6 and H-8 have been moved downfield only for 0.10 ppm in the acetylated product. But our ¹H-NMR result was the same as that previously described by Silva *et al.* 2009 and Scio *et al.* 2003 [9,10,19], for Rhamnocitrin and by Rossi *et al.*, for three acetylated compound [8]. These inferences and a detailed analysis of the ¹H-NMR, ¹H-¹H NOESY, HMQC, and HMBC spectrum and comparison with literature data suggested a Rhamnocitrin 3-sulphate.

Rhamnocitrin 3-sulphate was isolated as a yellow amorphous powder, (250 mg); UV λ max (MeOH) nm: 340, 265; ¹H NMR (DMSO; 300 MHz); 8.13 (2H, d, J = 8.9, H-2' and H-6'), 6.85 (2H, d, J=8.9, H-3' and H-5'), 6.71 (1H, s, H-8), 6.34 (1H, s, H-6), 3.86 (3H, s, OCH₃ -7). ¹³C NMR (DMSO; 75 MHz) 177.8 (C-4), 164.9 (C-7), 161.0 (C-4'), 160.0 (C-5), 156.6 (C-3), 156.0 (C-9), 132.5 (C-2), 130.8 (C-2' and C-6'), 121.1 (C-1'), 115.0 (C-3' and C-5'), 105.1 (C-10), 97.5 (C-8), 92.1 (C-6), 56.0 (OCH₃). MS (70 eV) m/z

$C_{16}H_{11}SO_9$; 379 (M-H), 315 (M-H-SO₂); 299 (M-H-SO₃), 284 (M-H-SO₃-CH₃); 271 (M-H-SO₃-C=O); 255 (M-H-SO₃-CH₃-HC=O); 243 (M-H-SO₃-CH₃-CH=C=O); 227 (M-H-SO₃-OCH₃-CH=C=O).

3,5,4'-Triacetoxy-7-methoxyflavone or **3,5,4'-Triacetoxy-Rahmnocitrin**. UV λ max (MeOH) nm: 304, 252; ¹H NMR (300 MHz, CDCl₃): 7.82 (d, J = 8.7, H-2', H-6'), 7.22 (d, J = 8.7, H-3', H-5'), 6.82 (d, J = 2.4, H-8), 6.43 (d, J = 2.4, H-6), 3.89 (s, OMe), 2.41 (s, OAc), 2.32 (s, OAc), 2.30 (s, OAc).

5. Conclusions

The study has demonstrated to be very important

particularly for the production scale of Rahmnocitrin 3-sulphate. Recrystallization is very simple, easy, cheap and perfect for isolation of pure Rahmnocitrin 3-sulphate from *Tetracera alnifolia* which could be considered as raw material for several usages.

ACKNOWLEDGEMENTS

This study is carried out under the program of Chimie Recherche et Développement Pharmaceutique (CHIRED-CONGO). The authors are deeply indebted to Madame Kouka of the CERVE for plant identification.

Annexes

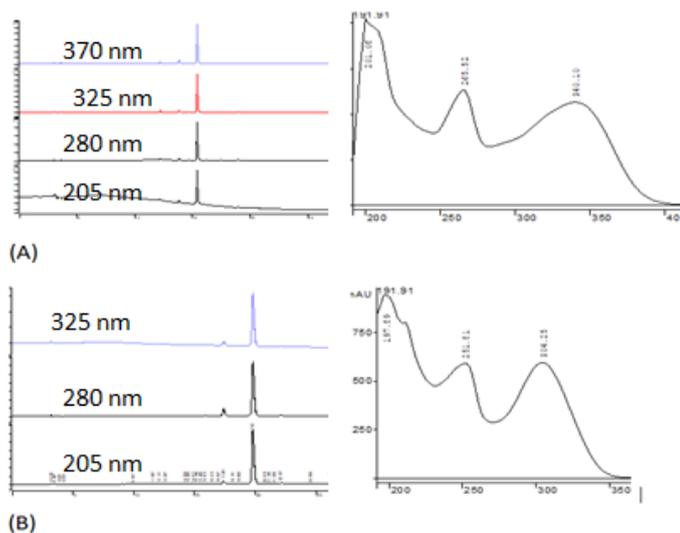
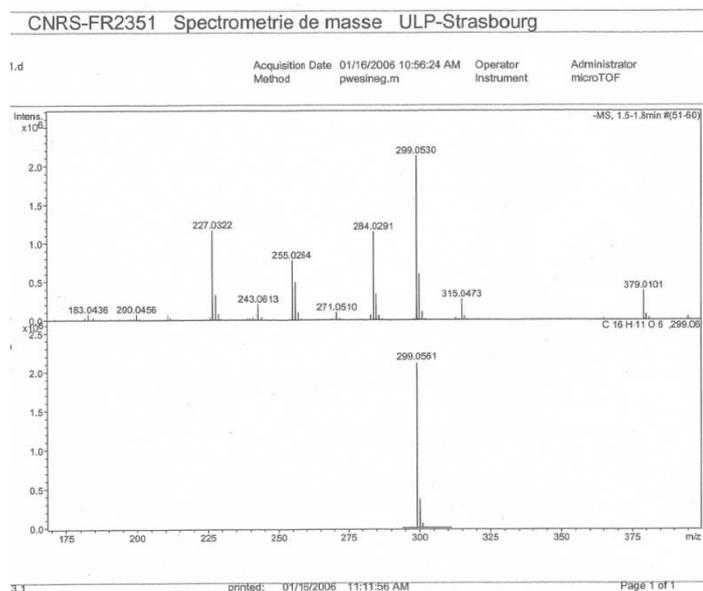
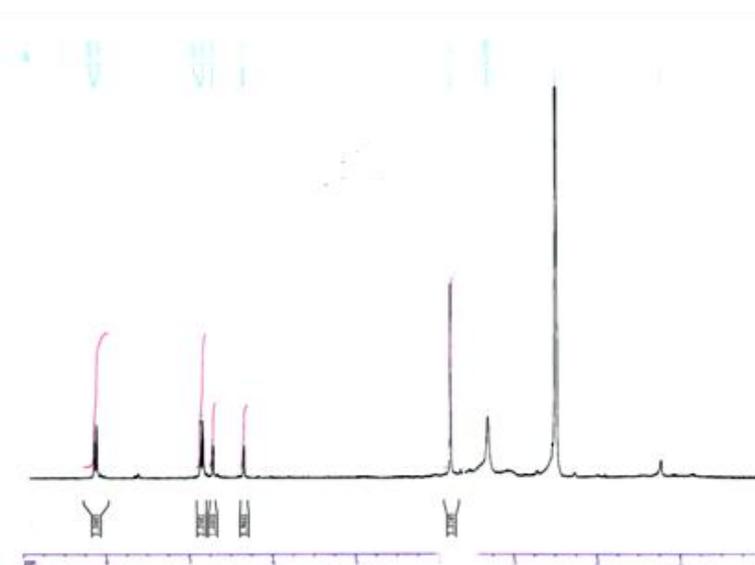
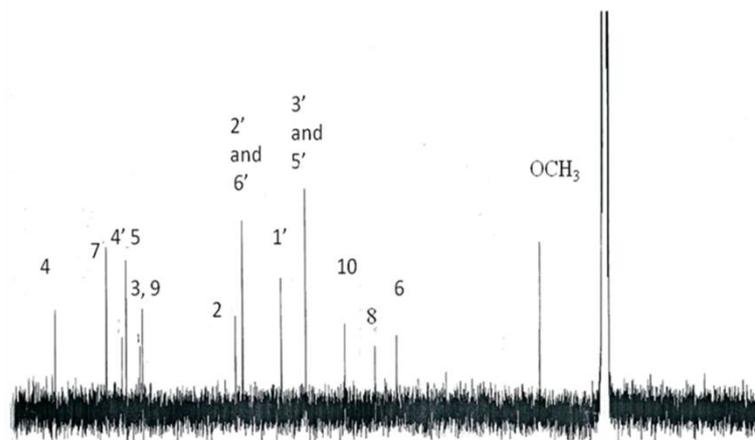
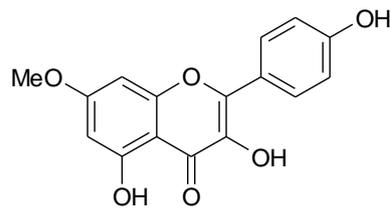
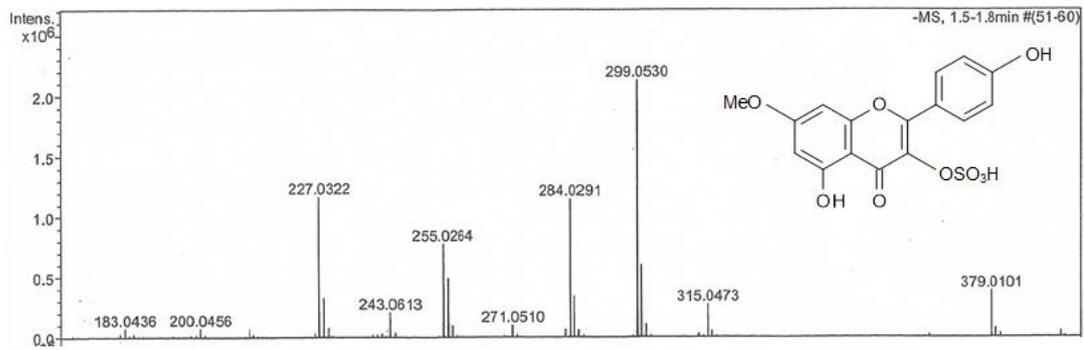


Figure 1. HPLC chromatograms and UV/Vis spectra of rhamnocitrin 3-sulphate (A) and rhamnocitrin triacetate (B). Analytical C18-HPLC: Pump Varian 9010; DAD UV detector, Varian Prostar 330, Column: Symmetry C18, 5 μ m, 4,6x250 mm, Rate of flow: 1 ml/min; Injection volume: 30 μ l. Monitored at 205, 280, 325 and 370 nm for (A); sample concentration 1mg/ml; eluent: acetonitrile/water mode gradient





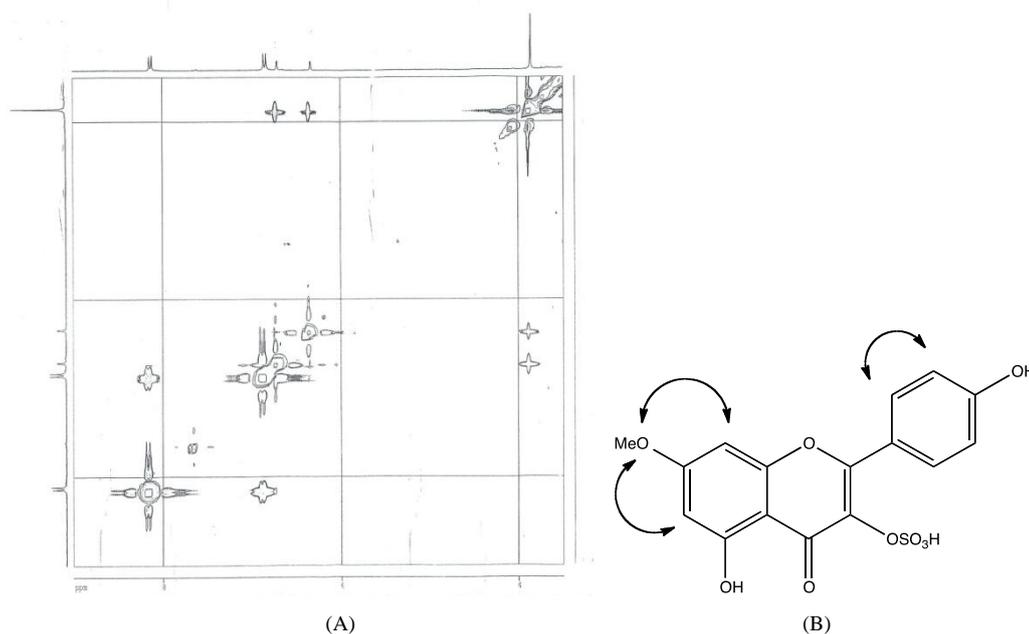


Figure 6. (A) ^1H - ^1H NOESY-NMR Spectra of Rhamnocitrin 3-sulphate, (B) nOe key NMR correlations of Rhamnocitrin 3-sulphate

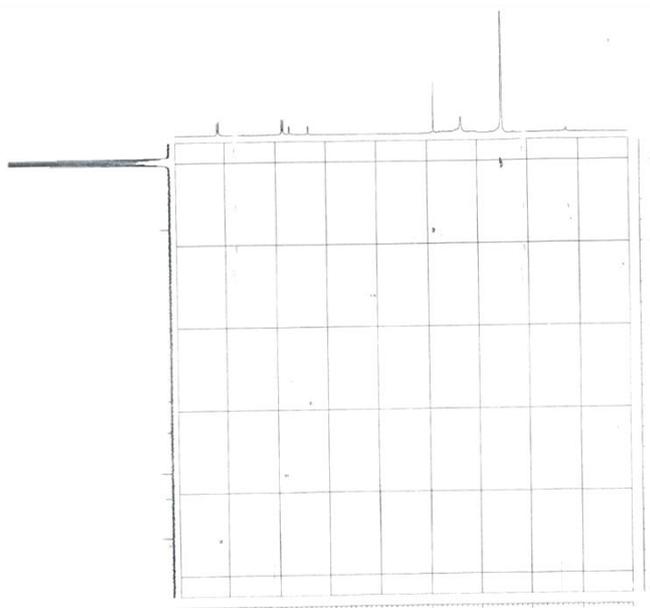


Figure 7. ^1H - ^{13}C HSQC NMR Spectra of Rhamnocitrin 3-sulphate

REFERENCES

- [1] Kosei Yamauchi, Tohru Mitsunaga, Irmanida Batubara. Isolation, Identification and Tyrosinate Inhibitory Activities of the Extractives from *Allamanda cathartica*. *Natural Resources*. 2011, 2: 167-172.
- [2] Chaabi Mehdi, Beghidja Nouredine, Benayache Samir, and Lobstein Annelise. Activity-Guided Isolation of Antioxidant Principles from *Limoniastrum feei* (Girard) Batt. *Z. Naturforsch.* 2008, 63c: 801-807.
- [3] Adjanohoun J. E., Ahyi M. R., Ake Assi L., Baniakina J., Chibon P., Cusset G., Doulou V., Enzanza A., Eyme J., Goudote E., Keita A., Mbemba C., Mollet J., Moutsambote J.-M., Mpati J., SITA P. *Médecine Traditionnelle et pharmacopée: contribution aux études ethnobotaniques et floristiques en République Populaire du Congo*. Editions A.C.C.T. 1988, Paris, 605p.
- [4] Adjanohoun J. E., Adjakidje V., Ahyi M. R. A., Ake Assi L., Akoegninou A., d'Almeida J., Apovo F., Boukef K., Chadare M., Cusset G., Dramane K., Eyme J., Gassita J.-N., Gbaguidi N., Goudote E., Guinko S., Houngnon P., Issa Lo, Keita A., Kiniffo H. V., Kone-Bamba D., Musampa Nseyya A., Saadou M., Sodogandji Th., de Souza S., Tchabi A., Zinsou Dossa C., Zohoun Th. *Médecine Traditionnelle et pharmacopée: contribution aux études ethnobotaniques et floristiques en République Populaire du Bénin*. Editions A.C.C.T. 1989, Paris, 227p.

- [5] Ndounga M., Mpati J., Chen Jian Min, Zhaou Yuan Peng, Bilala J.P., Sianard D.F., Koubemba M.C. Makambila. Etude préliminaire de l'activité antibactérienne de quelques plantes médicinales de la flore congolaise. *Revue Méd. Pharm. Afr.* 1991, 5(1): 33-42.
- [6] Gonnet J.-F. and Jay M., Les aglycones flavoniques d'*Anthyllis vulneraria*. *Phytochemistry*. 1972, 11(7): 2313-2316.
- [7] Lin C.N., Chung M.I., Gan K.H., Lu C.M. Flavonol and anthraquinone glycosides from *Rhamnus formosana*. *Phytochemistry*. 1991, 30: 3103-3106.
- [8] Stevens Jan F., Hart Henk and Wollenweber Eckhard. The Systematic and Evolutionary Significance of Exudate Flavonoids in *Aeonium*. *Phytochemistry*. 1995, 39(4): 805-813.
- [9] Rossi Maria Helena, Yoshida Massayoshi and Soares Maia Jose Guilherme. Neolignans, Styrylpyrones and Flavonoids from an *Aniba* Species. *Phytochemistry*. 1997, 45(6): 1263-1269.
- [10] Silva, T.M.S.; Carvalho, M.G.; Braz-Filho, R. Spectroscopy study on structural elucidation of flavonoids from *Solanum jabrense* Agra & NEE e *S. paludosum* Moric. *Quim. Nova*. 2009, 32(5): 1119-1128.
- [11] Martini N.D., Katerere D.R.P., Eloff J.N., Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae). *Journal of Ethnopharmacology*. 2004, 93: 207-212.
- [12] Okada Y., Miyauchi N., Suzuki K., Kobayashi T., Tsutsui C., Mayuzumi K., Nishibe S., Okuyama T. Search for naturally-occurring substances to prevent the complications of diabetes. 2. Inhibitory effect of coumarin and flavonoid derivatives on bovine lens aldose reductase and rabbit platelet-aggregation. *Chemical & Pharmaceutical Bulletin*. 1995, 43: 1385-1387.
- [13] Ram S.N., Dwivedi S.P.D., Pandey V.B., Rao Y.V. Pharmacological actions of kaemperol-7-methyl ether isolated from *Rhamnus triquerta*. *Fitoterapia*. 1989, 60: 273-274.
- [14] Gurni Albert A. and Kubitzki Klaus. Flavonoids chemistry and systematics of the Dilleniaceae. *Biochemical Systematics and Ecology*. 1981, 9(2/3): 109-114.
- [15] Gurni Albert A., König Wilfried A. and Kubitzki Klaus. Flavonoid glycosides and sulphates from the Dilleniaceae. *Phytochemistry*. 1981, 20(5): 1057-1059.
- [16] Bohlman F. and Zdero C. *Tetrahedron Letters*. 1967, 33: 3239.
- [17] Harborne Jeffrey B., King Linda. Flavonoid sulphates in the Umbelliferae. *Biochemical Systematics and Ecology*. 1976, 4(2): 111-115.
- [18] Scio Elita, Ribeiro Antônia, Tânia M.A. Alves, Alvaro J., Romanha c, Filho José Dias de Souza, Cordell Geoffrey A., Zani Carlos L.. Diterpenes from *Alomia myriadenia* (Asteraceae) with cytotoxic and trypanocidal activity. *Phytochemistry*. 2003, 64: 1125-1131.
- [19] March Raymond E., Miao Xiu-Sheng. A fragmentation study of Kaempferol using electrospray quadrupole time-of-flight mass spectrometry at high mass resolution. *International journal of Mass Spectrometry*. 2004, 231:157-157.
- [20] Pizzolatti Moacir Geraldo, Anildo Cunha Jr., Szpoganicz e Eliandra de Sousa Bruno. Flavonoids glycosides from leaves and flowers of *Bauhinia forficata* (Leguminosae). *Quim. Nova*. 2003, 26(4): 466-469.